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SYNTHESIS OF NEW CARBOCYCLIC OXETANOCIN ANALOGUES

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Abstract: Carbocyclic oxetanocin analogues containing 3-fluoro or 3-hydroxy cyclobutyl moieties and different natural bases, have been prepared from the olefinic precursors either by direct fluoro-iodination (AgF-I₂) or by DAST fluorination of the bromhydrin. The latter allowed the synthesis of the corresponding phosphonate derivatives as well.

INTRODUCTION

Oxetanocin A (1) is the first and so far unique example of a natural four-membered ring nucleoside. This compound has stimulated a great synthetic effort since its discovery by Shimada and co-workers in 1986.¹ In particular, the discovery in 1990^{2,3} that carbocyclic analogues of oxetanocin A 2 and 3 exhibit more potent antiherpes activities than 1, as well as anti-HIV activity, led numerous groups to investigate original syntheses of carbocyclic oxetanocin and related substances (Figure 1).⁴⁻⁷

Figure 1

Among these compounds, SQ-32,829 (4),8 5,9,10 fluoro-derivative 6,12 phosphonate derivative 7¹³ and cyclobutene 8¹¹ have been synthesized and some of them exhibit potent anti-viral activities (Figure 2).

This article describes our synthesis of the previously unknown 3'-fluoro-cyclobutyl derivatives of 9, 10 and 11 as well as the phosphonate derivatives 12, 13 and 14 (Figure 3).

Figure 3

RESULTS AND DISCUSSION

For this purpose, we started from olefin intermediates 15 and 16 synthesized previously. The latter were dihydroxylated in good yields with osmium tetroxide (OsO₄) leading to diols 5 and their isomers 17 and 18.9

Scheme 1

15

16

However, as observed recently, direct fluorination of 17 and 18 with diethylaminosulfur trifluoride (DAST) led to decomposition 14 of the starting materials; the same fluorination of the protected precursor 19 led also to extensive decomposition and to a low yield of the target derivative 20 (Scheme 2). This compound was detected by the presence of a doublet at 3.41 ppm with a characteristic coupling constant of 18.38 Hz for $^3J_{CH2-F}$ in the 1H NMR spectrum of the reaction mixture.

Scheme 2

We then focussed on bromohydrins 21 and 22 which were obtained in good yield with aqueous N-bromosuccinimide (NBS), 15 under phase transfer conditions and which advantageously replaced diols 17 or 18 by avoiding the protection-deprotection steps. Isomer 22 was fluorinated with DAST in CH₂Cl₂ at O°C. As expected 16 inversion of configuration was observed according to NMR data and led to 23. A byproduct 24, isolated in low yield, resulted from a competitive "intramolecular elimination reaction" of O=SFNEt₂ group and HF (Scheme 3). Accordingly, treatment of alcohols 21 or 22 with thionyl chloride or phosphorus oxychloride in pyridine afforded 24 in high yield. 12

Scheme 3

reagents and conditions: a : NBS (1.2 eq.)/KOH (1.2 eq.), AcOEt/H₂O, 15min., 97%ratio cis/trans = 55/45. b : DAST, CH₂Cl₂, 0°C, 15 min., 79% of 23. c : KOAc , cat. 18-crown-6, DMF, 25°C, 4 h; 95%. d: liq. NH₃- EtOH , 40°C, 24 h; 66%. e : HCl; H₂O; reflux; 2h; 91%.

Nucleophilic substitution of the bromine atom in 23 was then performed with potassium acetate in N,N-dimethylformamide (DMF) and treatment of resulting 25 with liquid ammonia led to adenine derivative 9

whereas chlorine hydrolysis in 25 afforded hypoxanthine derivative 11. However, the same methodology could not be used for the preparation of the guanyl derivative 10 owing to interference of DAST with the amino group of guanine precursor as observed previously in similar cases.^{17,18}

An alternative route for introduction of the 3'-fluoro moiety in the cyclobutane derivatives of adenine and guanine appeared to be the addition of iodine fluoride to the suitable 3'-methylene precursor (15, 16). This procedure was used by Moffatt and co-workers¹⁹ in the synthesis of Nucleocidin and, in the present case, did not require any protection of the amino group of the guanine precursor. Thus, fluoro iodination of 16 proceeded in good yield and the ratio of 26: 27 was always between 7/3 and 6/4 (Scheme 4).

This method has been also successfully carried out from 15. In each case the mixture of *cis-trans* isomers obtained was separated by column chromatography.

Scheme 4

reagents and conditions: a: I2; AgF; CH2Cl2; r.t.; 65 %. b: KOAc; DMF; 100°C; 5 hours. c: HCl 1N; H2O; 100°C;

Substitution of the iodine moiety in compound 26 was achieved by dry potassium acetate in DMF under the same conditions as substitution of bromine group in 23. This substitution turned out to be much easier than the substitution of iodine in the case of N^6 , N^6 -dibenzoyl-5'-deoxy-4'-fluoro-5'-iodo-2', 3'-o-isopropylidene adenosine 29^{19} where the resistance of the iodine function toward substitution was interpreted by the strong deactivating effect of an α fluorine atom due to its high electronegativity (Figure 4).

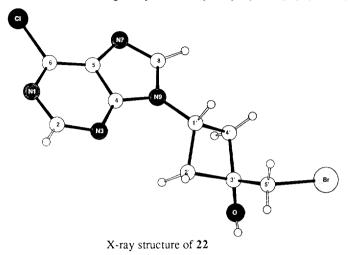
Figure 4

The present example illustrates that electronegative α -substituents exert a cumulative deactivating effect upon attempted displacement of iodine from 29 since fluorine by itself does not prevent nucleophilic substitution of the halogen atom in 23 or in 26. In addition, it should be mentioned here that catalytic amount of 18-crown-6

ether accelerated this substitution by a factor of four as shown by the substitution of bromine or iodine by acetate ion in 23 and 26, with or without crown ether, respectively.

Assignment of structure 22 was unambiguously resolved by x-ray crystallography. Its crystal structure

Figure 5



(Figure 5) shows the *cis* position of the hydroxyl moiety with respect to the base, and the *gauche* conformation of the CH₂Br substituent.

A summary of the ¹H NMR data concerning H-1 proton of the different couples of isomers is presented in Table 1. In the case of fluoro iodo cyclobutane derivative 27, a split quintet was observed for H-1 proton in the ¹H NMR spectrum in different solvents (DMSO-d₆, CDCl₃). This doubled quintet in 27 can be explained by a long-range W coupling (⁴J= 3Hz) of H-1 to the *trans* fluorine atom. This long-range coupling was not observed in the case of the *trans* fluoro isomer 26 where H-1 and F-3 are in *cis*. The quintet observed for H-1 in the latter case (26) as well as in the *trans* hydroxylated isomer 21 indicates equivalence of coupling constants of H-1 to both H-2 and H-4 protons. The H-1 signal in the *trans* hydroxylated isomers 21, 35 and *trans* fluoro isomer 26, always appeared as a quintet at lower field than in the *cis* hydroxylated isomers 22, 36 and *cis* fluoro isomer 27 which appeared as a more complex multiplet.

Table 1

Compound	21	22	26	27	35	36
H-1a	q	m	q	dq	q	m
δ	5.25	4.85	5.09	4.62	5.01	4.55
³ JH1-H2/4	8.50		8.32		8.42	
4 _{JH1-F}	-	-	-	3	-	-

 4 H-1 signals in the 1 H NMR spectra of compounds 21-36 in DMSO-d₆. q= quintet; m= multiplet; dq= doublet of quintet. δ : chemical shifts in ppm; J: coupling constants in Herz.

Furthermore assignment of the stereochemistry of the *trans* (35) and *cis* (36) bromohydrins is suggested by the comparison of their ¹H NMR spectra in DMSO-d6. Thus the hydroxyl group in 36 is oriented towards the

purine and therefore is subjected to a ring current with δ_{OH} of 5.77 ppm to be compared with δ_{OH} of 5.53 ppm in 35. The chemical shift of H-1 is also influenced by the proximity of OH in the *trans* isomer (35): δ = 5.01 ppm to be compared to a δ of 4.55 ppm in the *cis* isomer 36.

The Michaelis-Arbuzof reaction of bromohydrins 21 and 35, with triethylphosphite yielded the desired diethyl phosphonic acid ester derivatives 31 and 37, respectively. In the case of 6-chloropurine derivative 21, the reaction with triethylphosphite was performed at 100° C to avoid the displacement of the 6 chlorine atom of the purine by triethylphosphite (SNA_T reaction). The byproduct 32 was formed only in low yield (<10%) in these conditions, and in higher yield (>25%) above 120°C. Monoethylester 12-14, obtained by treatment with potassium or sodium hydroxide in aqueous dioxane, were purified by reversed phase HPLC (scheme 5).

Scheme 5

21
$$\xrightarrow{a}$$
 \xrightarrow{EtO}
 \xrightarrow{P}
 \xrightarrow{OEt}
 $\xrightarrow{NH_2}$
 $\xrightarrow{NH_2}$
 $\xrightarrow{NH_2}$
 $\xrightarrow{NH_2}$
 \xrightarrow{SI}
 \xrightarrow{OEt}
 $\xrightarrow{NH_2}$
 \xrightarrow{SI}
 \xrightarrow{OEt}
 $\xrightarrow{NH_2}$
 \xrightarrow{SI}
 \xrightarrow{OEt}
 $\xrightarrow{NH_2}$
 \xrightarrow{SI}
 \xrightarrow{SI}

reagents and conditions: a: (EtO)₃P; 100°C; 24 h; 85% of 31 and 8% of 32, or (EtO)₃P; reflux, 40% of 31, 25% of 32, a': (EtO)₃P; reflux; 24 h; 43% of 37; b: liq. NH₃; EtOH; 40°C; 24 h; 67%. c: NaOH (4 eq.); H₂O/dioxane; 65°C, 24 h. d: Na (4 eq.); MeOH; reflux; 90 min.; 93%. e: NBS (1.2 eq.); AcOEt/H₂O; 30 min.; 61%; f: KOH(4 eq.); H₂O/dioxane; 65°C, 24h.

These monoethylesters were synthesized as potential prodrugs, capable of being hydrolyzed in cells²¹ to the corresponding free phosphonic acids. These phosphonate derivatives (12-14) were considered as analogues of the expected monophosphate metabolites of 5a and of its guanine analogue. Antiviral activities were measured in primary peripheral blood mononuclear cells (PBMC) infected with HIV-1 (III B strain) or HIV-2 (D194 strain).²⁴ On these viruses, the most active compound 9 had respectively an IC50 of 3.9 μ M and 1.5 μ M, and compound 10 95 μ M and 46 μ M, whereas 30% inhibition of HIV-1 replication was observed with 11 at 100 μ M and 50% inhibition of HIV-2 at 45 μ M. No cytotoxicity was detected with compounds 9-11 up to 100 μ M (MTT assay). Phosphonate derivatives 12, 13 and 14 were found inactive on HIV-1 replication but slightly inhibited cell metabolism (10-20% at the highest concentration tested: 10-100 μ M).

CONCLUSION

This paper describes synthetic approaches toward 3'-fluorocyclobutyl nucleoside analogues. 20 The best general method for introducing a fluorine atom in the presence of an amino-purine (adenine or guanine) turned out to be the direct fluoro-iodination of an olefinic precursor by iodine fluoride generated *in situ* from silver fluoride and iodine, leading to a good yield of separable isomers. Contrary to the generally observed instability of 4'-fluoro-ribonucleosides 19 and as expected for carbocyclic nucleosides, the presence of a fluorine atom in the cyclobutane portion did not lead to the release of the purine under acidic conditions. This was shown, for example, by the high yield of guanine derivative 10 obtained from precursor 28 under acidic conditions. The nucleophilic substitution of iodine and bromine in the presence of an α fluoro group proved easy in the cyclobutyl derivatives studied.

To the best of our knowledge, we report in this article the first example of a Michaelis-Arbuzof reaction of a 6-chloropurine bromohydrin derivative with triethylphosphite.

EXPERIMENTAL

The melting points were taken on a Kofler hot stage apparatus and were uncorrected. Elemental analyses were performed by the "Service de Microanalyse", CNRS, ICSN, 91198 Gir sur Yvette, France. Fast atom bombardment (FAB) and chemical ionization (CI) mass spectra (MS) were obtained from the "Laboratoire de Spectrometrie de Masse", CNRS, ICSN, 91198 Gif sur Yvette, France. Proton nuclear magnetic resonance (¹H-NMR) spectra were recorded at 200 MHz on a Bruker AC200 spectrometer. Chemical shifts (δ) are reported in ppm units with tetramethylsilane as an internal standard and coupling constants (J) are given in hertz (Hz). For the sake of clarity, the same numbering has been used for the description of all NMR spectra, as follows:

Preparative HPLC of phosphonate monoester derivatives were performed on a Gilson equipment (305 pump with UV detection UV 115). A reverse phase C18 Dynamax (21.4mm x 25cm) colomn was used with ethanol-water 9:1 as eluent.

1-Bromomethyl-3-(6-Chloro-9H-purin-9-yl)-cyclobutanol 21 and 22. A solution of 15⁹ (1 g, 4.53 mmol) in AcOEt (20ml) was added to an aqueous solution (25ml) of KOH (380 mg, 6.8 mmol) and NBS

(1.05 g, 5.90mmol) and stirred vigorously for 15 min. The organic layer was dried over Na₂SO₄ and the volatile removed under reduced pressure to give a mixture of *cis* and *trans* alcohol isomers which were separated by colomn chromatography on silica gel (CH₂Cl₂-MeOH 95:5). *Cis* isomer **22** (768 mg, 53%): mp 134°C (amorphous solid). 1 H NMR (CDCl₃) δ : 8.7 (s, 1H, H-2), 8.2 (s, 1H, H-8), 4.84 (m, 1H, H-1), 3.85 (s, 2H, CH₂Br), 3.1 (m, 2H, cyclobutane), 2.85 (m, 2H, cyclobutane). M.S. (FAB): 317-319-321 (MH)⁺, 237-239 (MH-Br)⁺, 155-157 (6-chloropurine)⁺, 131. *Anal.* Calcd for C₁₀H₁₀BrClN₄O: C, 37.79; H, 3.15; N, 17.64; O, 5.04. Found: C, 38.05; H, 3.13; N, 17.48; O, 5.06. *Trans* isomer **21** (628 mg, 44%): mp 128°C (amorphous solid). 1 H NMR (CDCl₃) δ : 8.7 (s, 1H, H-2), 8.05 (s, 1H, H-8), 5.25 (q, 1H, J=8.5 Hz, H-1), 3.65 (s, 2H, CH₂Br), 3.1 (m, 2H, cyclobutane), 2.8 (m, 2H, cyclobutane). M.S. (CI, isobutane): 317-319 (MH)⁺, 237-239 (MH-Br)⁺, 181-183 (MH-BrCH₂COCH₃)⁺, 155-157 (6-chloropurine)⁺. *Anal.* Calcd for C₁₀H₁₀BrClN₄O: C, 37.79; H, 3.15; N, 17.64; O, 5.04. Found: C, 38.04; H, 3.23; N, 17.73; O, 5.09.

6-Chloro-9H-9-(cis-3-bromomethyl-trans-3-fluoro-cyclobutyl) purine 23. To a stirred solution of 22 (1.483 g, 4.47 mmol) in cooled CH₂Cl₂ (50 ml, 0°C) was slowly added DAST (0.74 ml, 5.6 mmol). After 15 min, the reaction was quenched by pouring into a saturated and cooled aqueous solution of K₂CO₃ (50 ml). The organic layer was dried over Na₂SO₄ and the volatile removed under reduced pressure to give a residual oil which was purified by chromatography on silica gel column (CH₂Cl₂-MeOH 95:5). *Trans*-fluoro isomer 23 was eluted first and was always contaminated with a small amount of the elimination product 24. This mixture was used without further purification in the next step. (yield 79%); mp 107°C. ¹H NMR (CDCl₃) δ: 8.75 (s, 1H, H-2), 8.15 (s, 1H, H-8), 5.25 (q, 1H, J=8.5Hz, H-1), 3.85 (d, 2H, J=21.5 Hz, CH₂Br), 3.25 (m, 2H, cyclobutane), 3 (m, 2H, cyclobutane). M.S. (CI, isobutane): 319-321 (MH)+, 299-301 (MH-HF)+, 181-183 (MH-BrCH2COCH3)+, 155-157 (6-chloropurine)+. A second compound (24) was eluted as an oil after evaporation and crystallized from cyclohexane: (65 mg,6%); mp 112-114°C. ¹H NMR (CDCl₃) δ: 8.77 (s, 1H, H-2), 8.24 (s, 1H, H-8), 6.15 (q, 1H, J=3Hz, vinylic H), 5.20 (q, 1H, J=8.5Hz, H-1), 3.44 (m, 2H, cyclobutane), 3.40 (m, 2H, cyclobutane). M.S. (CI, isobutane): 299-301 (MH)+, 255-257, 221-223, 155-157 (6-chloropurine)+. *Anal.* Calcd for C₁₀H₈BrClN₄: C, 40.10; H, 2.69; N, 18.70. Found: C, 39.94; H, 3.03; N, 18.73.

Acetic acid-cis-3-(6-Chloropurin-9-yl)-trans-1-fluoro-cyclobutyl methylester 25. A stirred solution of 23 (450 mg, 1.4 mmol) in DMF (20 ml) was treated with dry KOAc (548 mg, 5.6 mmol) and catalytic 18-crown-6 (10 mg). After total consumption of the substrat (4 hours), the reaction was quenched by pouring into iced water (50 ml) and the solution was extracted with CH₂Cl₂ (2x50 ml). The organic layer was dried over Na₂SO₄ and the volatile removed under reduced pressure to give a residual oil which was purified by chromatography on silica gel column (AcOEt-heptane 2:1). *Trans* fluoro isomer 25 (400 mg, 95%) was obtained as a colourless oil. ¹H NMR (CDCl₃) δ : 8.7 (s, 1H, H-2), 8.1 (s, 1H, H-8), 5.3 (q, 1H, J=8.5Hz, H-1), 4.4 (d, 2H, J=21.5Hz, CH₂OAc) 2.8-3.4 (m, 4H, cyclobutane), 2.1 (s, 3H, CH₃). M.S. (CI, isobutane): 299-301 (MH)⁺, 155-157 (6-chloropurine)⁺. *Anal*. Calcd for C₁₂H₁₂CIFN₄O₂: C, 48.24; H, 4.02; N, 18.76. Found: C, 48.38; H, 4.15; N, 18.64.

trans-1-fluoro-cis-3-(adenin-9-yl)-cyclobutyl methanol 9. A 150 ml stainless steel autoclave was charged with a solution of 25 (192 mg, 0.64 mmol) in 50 ml ethanol/liquid ammonia (3/1) and heated at 40°C for 24 hours. After cooling to room temperature and removal of the solvent, the residual oil was purified by colomn chromatography on silica gel (CH₂Cl₂-EtOH 9:1). trans -fluoro isomer 9 was obtained in 66% yield as an amorphous solid: mp 168°C. 1 H NMR (DMSO d₆) δ : 8.3 (s, 1H, H-2), 8.17 (s, 1H, H-8), 7.27 (s, 2H, NH₂), 5.25 (q, 1H, J=8.5Hz, H-1), 5.2 (t, 1H, J'=4.5Hz, OH), 3.7 (dd, 2H, J=21.5Hz and J'=4.5Hz,

<u>CH2</u>OH), 2.65-3.1 (m, 4H, cyclobutane). M.S. (EI): 238 (MH)⁺, 237 (M)⁺, 162 (MH-HOCH₂CF=CH₂)⁺, 135 (adenine)⁺. Anal. Calcd for $C_{10}H_{12}FN_5O$: C, 50.63; H, 5.1; N, 29.52. Found: C, 50.6; H, 5.19; N, 29.63.

Cis-9-(trans-3-fluoro-cis-3-hydroxymethyl-cyclobutyl) hypoxanthine 11. A solution of 25 (367 g, 1.22 mmol) in aqueous 1N HCl (10 ml) was refluxed for 2 hours. After cooling to room temperature, the solution was evaporated under reduced pressure to give a residual oil which was purified by colomn chromatography on alumina (type T) (AcOEt-EtOH 8:2). trans-fluoro isomer 11 was obtained in 91% yield: mp 245°C (amorphous solid). ¹H NMR (DMSO d₆) δ : 12.35 (broad s, 1H, NH), 8.3 (s, 1H, H-2), 8.1 (s, 1H, H-8), 5.25 (m, 1H, H-1), 3.65 (dxd, 2H, J=21.5Hz and J'=4.5Hz, CH2OH), 2.6-3.05 (m, 4H,, cyclobutane). M.S. (EI): 238 (M)+, 162 (MH-HOCH₂CF=CH₂)+, 137 (hypoxanthine+H)+, 136 (hypoxanthine)+. Anal. Calcd for C₁₀H₁₁FN₄O₂, 1/2 H₂O: C, 48.58; H, 4.85; N, 22.67. Found: C, 48.7; H, 4.96; N, 22.59.

6-Chloro-9-(3-fluoro-3-iodomethyl-cyclobutyl)-9H-purin-2-yl amine 26 and 27. A solution of olefin **16**⁹(500 mg, 2.12 mmol) in dichloromethane was treated with AgF and iodine as described¹⁹ to afford a mixture of *cis* and *trans* isomers which were separated by column chromatography (silica gel; 2x 95 cm) eluting with ethyle acetate-heptane 2:1. *trans* Isomer **26** was eluted first (fluorine atom being in *trans* with respect to the base) 347 mg; mp: 200-202°C (ethanol): MS (CI, isobutane): 382. ¹H NMR (DMSO-d₆) δ: 8.26 (s, 1H, H-8), 6.90 (s, 2H, NH₂), 5.09 (q, 1H, J=8.32 Hz, H-1), 3.83 (d, 2H, I<u>CH₂</u>, 3 J_{H-F}= 24.23 Hz), 3.10-2.68 (m, 4H, cyclobutane). *Anal.* Calcd for C₁₀H₁₀N₅OCIFI: C, 31.48; H, 2.64; N, 18.35; F, 4.98. Found: C, 31.70; H, 2.68; N, 18.38; F,5.29. A second product was then obtained which corresponded to the *cis* isomer **27**: 172 mg (total yield: 65.3 %); mp: 202-204°C (ethanol); MS (CI, isobutane): 382; ¹H NMR (DMSO-d₆) δ: 8.25 (s,1H, H-8), 6.82 (s, 2H, NH₂), 4.62 (dq, 1H, H-1, 3 J=8.25 Hz; 4 J_{H-F}= 3Hz), 3.78 (d, 2H, I<u>CH₂</u>, 3 J_{H-F}= 24.26 Hz), 3.32-2.80 (m, 4H, cyclobutane). *Anal.* Calcd for C₁₀H₁₀N₅OClFI: C, 31.48; H, 2.64; N, 18.35; F, 4.98. Found: C, 31.77; H, 2.72; N, 18.07; F, 4.79.

Acetic acid-3-(2-amino-6-chloro-purin-9-yl)-trans-1-fluoro-cyclobutyl methylester 28. A solution of trans 26 (313.3 mg, 1 mmol) in dry DMF (100 ml) was treated with KOAc (804 mg, 8.2 mmol) at 100° C for 15 h with stirring. The mixture was then cooled, evaporated to dryness (oil pump) and the residue was partionned between dichloromethane (200 ml) and water (50 ml). The organic phase was washed twice with water (20 ml), dried (MgSO₄), filtered and evaporated to dryness. Purification was then carried out by column chromatography (2 x 95 cm) on silica gel prepared in AcOEt-heptane 1:1. The column was eluted successively with 1 L of AcOEt-heptane 1:1 and 1 L of AcOEt-heptane 2:1 to afford 28 as an oil which crystallized on standing. Yield: 79%; mp: 170-172°C (AcOEt). ¹H NMR (CDCl₃) δ : 7.73 (s, 1H, H-8), 5.43 (s, 2H, NH₂), 5.12 (q, 1H, J=8.38 Hz, H-1), 4.64 (d, 2H, AcOCH₂, ³J CH₂-F = 22.94 Hz), 3.27-2.80 (m, 4H, cyclobutane), 2.15 (s, 3H, CH₃). Anal. Calcd for C₁₂H₁₃N₅O₂CIF: C, 45.94; H, 4.18; N, 22.32; Cl, 11.48. Found: C, 45.95; H, 4.12; N, 22.42; Cl, 11.48.

2-Amino-9-(*trans*-3-fluoro-*cis*-3-hydroxymethyl-cyclobutyl)-1,9-dihydropurin-6-one **10.** A solution of acetoxy derivative **28** (100 mg, 0.318 mmol) in 1N HCl (100 ml) was heated at 100°C for 6 h, cooled and evaporated to dryness. The solid residue was dissolved in 2 ml of water and neutralized with 6N NaOH. A precipitate was obtained which was filtered and washed with cooled water. Yield: 96%; mp: > 260°C; ¹H NMR (DMSO-d₆) δ : 10.59 (s, 1H, NH), 8.03 (s, 1H, H-8), 6.42 (s, 2H, NH₂), 5.12-4.98 (m, 2H, OH, H-1), 3.64 (dd, 2H, CH₂OH, 3 J_{H-OH} = 4.75 Hz, 3 J_{H-F} = 20.81 Hz), 2.97-2.62 (m, 4H, cyclobutane). *Anal.* Calcd for C₁₀H₁₂N₅O₂F/1/3H₂O: C, 46.33; H, 4.92; N, 27.01; Found: C, 46.58; H, 4.95; N, 27.21.

6-Methoxy-9-(3-methylencyclobutyl)-9H-purin-2-yl amine 34. A solution of 6-chloro-9-(3-methylenecyclobutyl)-9H-purin-2-yl amine 16^9 (4.5 g; 19.1 mmol) and sodium (1.2 g; 52 mmol) in methanol

(400 ml) was heated under reflux for 90 min. After evaporation to dryness, the residue was dissolved on CH_2Cl_2 (200 ml) and washed with water (4 x 50 ml). The organic phase was dried (MgSO₄), filtered and evaporated to give an oil which gave crystals on trituration with ether (4.1 g; 93 %). mp = 123-124°C. ^{1}H NMR (DMSO-d₆) δ 8.10 (s, 1H, H-8), 6.45 (s, 2H, NH₂), 4.98 (m, 2H, CH₂=); 4.93 (q, 1H, J=8Hz, H-1), 3.99 (s, 3H, OCH₃), 3.39 (m, 2H, cyclobutane), 3.15 (m, 2H, cyclobutane). *Anal.* Calcd for $C_{11}H_{13}N_5O$: C, 57.29; H, 5.68; N, 30.38. Found: C, 57.14; H, 5.62; N, 30.30.

1-Bromomethyl-3-(2-amino-6-methoxy-9H-purin-9-yl) cyclobutanol 35 and 36. A solution of 34 (1.7 g; 7.35 mmol) in AcOEt (225 ml) was stirred vigorously with H₂O (375 ml) and NBS (1.56 g; 8.83 mmol) at room temperature for 30 min. The organic phase was separated while the aqueous phase was extracted 3 times with AcOEt (50 ml). The combined organic phases were washed twice with 5 % aqueous NaHSO₃ (30 ml portions) and twice with water (30 ml portions), dried (MgSO₄), filtered and evaporated to dryness. The oily residue was adsorbed on silica gel and subjected to column chromatography (50 x 2.5 cm), eluting with CH₂Cl₂-EtOH 95:5. A first compound was eluted (36) (308 mg) mp = 170-172°C (CH₂Cl₂); Rf = 0.33. ¹H NMR DMSOd₆ δ : 8.03 (s, 1H, H-8), 6.38 (s, 2H, NH₂), 5.77 (s, 1H, OH), 4.55 (m, 1H, H-1), 3.95 (s, 3H, OCH₃), 3.74 (s, 2H, CH₂ Br), 2.80-2.50 (m, 4H, 2 x CH₂). *Anal.* Calcd for C₁₁H₁₄N₅O₂ Br C, 40,26; H, 4.30; N, 21.34. Found : C, 40.13; H, 4.29; N, 21.15. A second compound (35) was eluted (1153 mg) Rf = 0.24; mp = 198-202°C (CH₂Cl₂) (decomposition) ¹H NMR DMSO-d₆ δ : 8.05 (s, 1H, H-8), 6.40 (s, 2H, NH₂), 5.53 (s, 1H, OH), 5.01 (q, 1H, J=8.42 Hz, H-1), 3.95 (s,3H, CH₃), 3.77 (s, 2H, CH₂ Br), 2.68 (m, 2H, cyclobutane), 2.42 (m, 2H, cyclobutane). *Anal.* Calcd for C₁₁H₁₄N₅O₂Br: C, 40.26; H, 4.30; N, 21.34. Found : C, 40.44; H, 4.30; N, 21.09.

trans-1-Diethoxyphosphonomethyl-trans-3-(2-amino-6-methoxy-9H-purin-9-yl)-

cyclobutanol 37. A solution of bromo derivative **35** (753 mg; 2.29 mmol) and freshly distilled triethyl phosphite (250 ml) was heated under N₂ at 120°C for 24 h.Excess of triethylphosphite was evaporated *in vacuo* (oil pump) and the residue was purified by column chromatography on silica gel prepared in CH₂Cl₂. Elution was carried out successively with CH₂Cl₂, CH₂Cl₂-EtOH 99:1 and 98:2 to eliminate some impurities. Compound **37** was then eluted with CH₂Cl₂-EtOH 97:3 to give a solid which was washed with ether. Yield: 43.5% (385 mg); mp 165°C. ¹H NMR (CD₃OD) δ: 8.24 (s, 1H, H-8), 5.62 (s, broad, 3H, NH₂, OH), 5.25 (q, 1H, H-1, J=8.4Hz), 4.14 (m, 7H, OCH₃,2x<u>CH₂OP</u>), 3.01-2.75 (m, 4H, 2xCH₂), 2.47 (d, 2H, CH₂-P, J=17.10 Hz), 1.35 (t, 6H, 2xCH₃). *Anal.* Calcd for C₁₅H₂₄N₅O₅ P: C, 46.75; H, 6.28; N, 18.17. Found: C, 46.93; H, 6.24; N, 17.92.

trans-1-Diethoxyphosphonomethyl-*trans*-3-(6-chloro-9H-purin-9-yl)-cyclobutanol 31. A solution of 21 (600 mg, 1.89 mmol) in triethylphophite (5 ml) was heated at 100°C for 24 h. The solution was then evaporated under reduced pressure to give a residual oil which was purified by colomn chromatography on silica gel (CH₂Cl₂-MeOH 92:8). *Cis* phosphonate 31 was obtained as an amorphous solid (601 mg, 85%); mp 96°C; ¹H NMR (CDCl₃) δ: 8.65 (s, 1H, H-2), 8.1 (s, 1H, H-8), 5.25 (q, 1H, J=8.5Hz, H-1), 4.1 (m, 4H, CH₃CH₂), 3.1 (m, 2H, cyclobutane), 2.8 (m, 2H, cyclobutane), 2.4 (d, 2H, CH₂P, J=27.5Hz), 1.3 (t, 6H, CH₃CH₂ClN₄O₄P, 2/3 H₂O: C, 43.49; H, 5.56; N, 14.49. Found: C, 43.81; H, 5.31; N, 14.11. A second compound (32) was obtained as an oil in 8% yield: ¹H NMR (CDCl₃) δ: 9.1 (s, 1H, H-2), 8.2 (s, 1H, H-8), 5.3 (q, 1H, J=8.5Hz, H-1), 4.35 (m, 4H, 2xCH₂CH₃), 4.15 (q, 4H, J=8.5Hz, 2xCH₂CH₃), 3.1 (m, 2H, cyclobutane), 2.75 (m, 2H, cyclobutane), 2.4 (d, 2H, J=27.5Hz, CH₂P), 1.4 (t, 6H, J=8.5Hz, CH₂CH₃), 1.3 (t, 6H, J=8.5Hz, CH₂CH₃). M.S. (FAB): 499 (M+Na)⁺, 477 (MH)⁺, 257 (Et₂O₃P-purine)⁺, 201, 123.

trans-1-Diethoxyphosphonomethyl-trans-3-(6-amino-9H-purin-9-yl)-cyclobutanol 33. A 150 ml stainless steel autoclave was charged with a solution of 31 (300 mg, 0.8 mmol) in 50 ml of ethanol-liquid ammonia (3:1) and heated at 40° C for 24 hours. After cooling to room temperature and removal of the solvent, the residual oil was purified by chromatography on silica gel column (CH₂Cl₂-EtOH 8:2) to give 33 as an oil (191 mg, 67%); ¹H NMR (CD₃OD) 8: 8.25 (s, 1H, H-2), 8.15 (s, 1H, H-8), 5.25 (m, 1H, H-1), 4.6 (broad s, 3H, OH and NH₂), 4.15 (m, 4H, $2\times CH_2$ CH₃), 3.05 (m, 2H, cyclobutane), 2.75 (m, 2H, cyclobutane), 2.47 (d, 2H, J=27.5Hz, CH_2 P), 1.3 (t, 6H, J=8.5Hz, $2\times CH_3$). M.S. (CI, isobutane): 356 (MH)⁺, 136 (adenine)⁺. Anal. Calcd for C₁₄H₂₀ClN₄O₄P, H₂O: C, 45.04; H, 6.48; N, 18.76. Found: C, 44.93; H, 6.31; N, 18.31.

General procedure for the synthesis of phosphonate potassium (or sodium) salt monoethylesters. A solution of phosphonate diethylester 31, 33 or 37 (0.3 mmol) in distilled dioxane (10 ml) and 4N aqueous KOH or NaOH (10 ml) was stirred at 75° C for 6h. The mixture was cooled, neutralized with concentrated aqueous HCl and evaporated to dryness. The residue was extracted with absolute EtOH and subjected twice to reverse phase chromatography eluting with ethanol-water 9:1.

2-Amino-9-(cis-3-ethoxyphosphonomethyl-trans-3-hydroxycyclobutyl)-1,9-dihydropurin-6-one potassium salt 13. Following the general procedure from 37 in 4N aqueous KOH, 13 was obtained as a hygroscopic solid in 42% yield; mp 202-204 °C. 1 H NMR (CD3OD) δ : 7.88 (s, 1H, H-8), 5.05 (q, 1H, J=8.5 Hz, H-1), 4.89 (broad s, OH, NH₂, NH), 3.95 (m,2H, OCH2CH3), 2.92 (m, 2H, cyclobutane), 2.63 (m, 2H, cyclobutane), 2.15 (d, 2H, CH2-P, 2 J=16.6Hz), 1.25 (t, 3H, J=7.0 Hz, CH3CH2). *Anal.* Calcd for C₁₂H₁₇N₅O₅PK, 3 H₂O: C, 33.10; H, 5.32; N, 16.08. Found: C, 33.03; H, 5.39; N, 15.98.

trans-1-Ethoxyphosphonomethyl-*trans*-3-(6-amino-9H-purin-9-yl)-cyclobutanol sodium salt 12. Following the general procedure from 33 in 4N aqueous NaOH, 12 was obtained as a solid in 70% yield; mp 196-202°C; ¹H NMR (CD₃OD) δ: 8.33 (s, 1H, H-2), 8.19 (s, 1H, H-8), 5.23 (q, 1H, H-1, J=8.5 Hz), 3.94 (q, 2H,J= 7 Hz, <u>CH₂CH₃</u>), 2.97 (m, 2H, H-3,H-4), 2.70 (m, 2H, H-3, H-4), 2.12 (d, 2H, J=16.7 Hz, CH₂-P), 1.25 (t, 3H, CH₂<u>CH₃</u>, J=7.0 Hz). *Anal*. Calcd for C₁₂H₁₇N₅O₄PNa,2 H₂O: C, 39.78; H, 5.84; N, 19.33. Found: C, 39.34; H, 5.51; N, 19.61.

9-(cis-3-Ethoxyphosphonomethyl-trans-3-hydroxycyclobutyl)-1,9-dihydro-purin-6-one sodium salt 14. Following the general procedure from 31 in 4N aqueous NaOH, 14 was obtained as an oil in 38% yield. H NMR (CD3OD) δ : 8.55 (s,1H,H-2), 8.19 (s, 1H, NH), 8.03 (s,1H,H-8),5.26(q, 1H,H-1, J=8.4 Hz), 3.94 (m, 2H, CH2CH3), 2.90 (m, 2H, H-2, H-4), 2.60 (m, 2H, H-2, H-4), 1.25 (t,3H, CH2CH3, J=4.2Hz). Anal. Calcd for $C_{12}H_{16}N_4O_5PNa$, 2 H2O: C, 39.67; H, 5.55; N, 15.42; Found: C, 40.01; H, 5.42; N, 15.28.

X-Ray structure determination of compound 22. The main crystal data are summarized as follows: $C_{10}H_{10}BrClN_4O$, M=317.6. Crystal dimensions $0.6 \times 0.4 \times 0.3$ mm³. Automatic, graphite monochromated ($\lambda=1.5418$ Å) 4-circle Nonius diffractometer. Monoclinic, $P2_1/n$, Z=4. a=7.047 (3), b=8.983 (7), c=19.295 (5) Å and $\beta=91.86$ (2)°. V=1221 (1) Å³, $d_x=1.71$, $\mu=6.6$ cm⁻¹. $\omega/2\theta$ scan mode (20 < 120°), lhl < 7, k < 10, I < 21 gave 2039 reflexions in which 1674 were independent and > 3 σ (I). Lorentz-polarization, no absorption corrections. Direct methods²² and full-matrix least squares²³: C, N, O, Br, Cl anisotropic, H isotropic refinement to R=5.4%. Weighting scheme $w=[\sigma^2(F)+0.0002 F^2]^{-1}$, σ from counting statistics. Supplementary material is available on request.

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